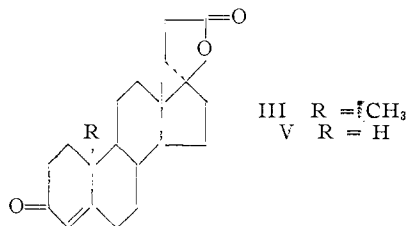


with metallic lithium⁴ thereby causing reduction of both the triple bond and the aromatic A ring. Hydrolysis with mineral acid followed by chromatography yielded 3-(3-oxo-17 β -hydroxy-19-nor-4-androsten-17 α -yl)propionic acid γ -lactone (V), m.p. 126.5–127° (137–138°), $[\alpha]_D^{25} +22.7^\circ$ (CHCl₃); *Anal.* Calcd. for C₂₁H₂₈O₃: C, 76.79; H, 8.59. Found: C, 76.49; H, 8.34.



Bioassay⁵—reversal of the effect of desoxycorticosterone (DOC) on the urinary Na–K ratio of adrenalectomized rats was used as a measure of potency. If progesterone, recently described as capable of reversing the effect of DOC,⁶ is assigned an activity of 1, then III and V were found to have an activity of approximately 8 and 27, respectively. In similar tests III and V also reversed the electrolyte effects of aldosterone.

(4) A. L. Wilds and N. A. Nelson, *THIS JOURNAL*, **75**, 5360 (1953).

(5) The details of bioassay will be reported elsewhere.

(6) R. L. Landau, D. M. Bergenstal, K. Lugibihl, and M. E. Kasch, *J. Clin. Endocrinol. and Metabolism*, **15**, 1194 (1955).

DIVISIONS OF CHEMICAL AND BIOLOGICAL RESEARCH
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RECEIVED JULY 24, 1957

CYCLIZATION IN GAMMA RAY IRRADIATED HIGH DENSITY POLYETHYLENE

Sir:

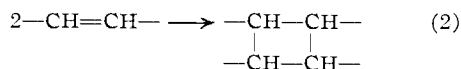
Material balance in the irradiation of polyethylene has never been convincingly attained, especially at liquid nitrogen temperatures where $G(\text{H}_2)$ and $G(-\text{CH}=\text{CH}-)$ are essentially equal to their values at room temperature while cross link formation is sharply reduced.¹

As adapted to the high density polyethylene, Marlex-50, and considering no chain scission, the material balance equation can be written

$$[\text{H}_2] + [\text{Vi}^0 - \text{Vi}] = \frac{1}{2}[\text{R}] + [\text{VI}] + [\text{C.L.}] + [\text{E.L.}] + [\text{S.L.}] + [\text{R.L.}] \quad (1)$$

where brackets signify moles per gram of substances produced during the irradiation except for $\text{Vi}^0 - \text{Vi}$ which represents moles/g. of vinyl groups destroyed. R, VI, C.L., E.L., S.L. and R.L. signify free radicals, vinylene groups, cross, end,² square and ring links, respectively.

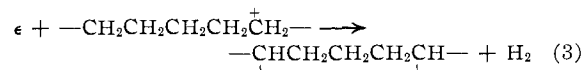
By "square" links we mean double cross links formed by a reaction such as



(1) A. Charlesby and W. H. T. Davison, *Chem. and Ind.*, 232 (1957).

(2) A. Charlesby has proposed the term "end link," *Proc. Roy. Soc.*, **A231**, 521 (1955).

Such a square link is assumed in Eq. (1) to constitute one cross link as well as one square link. By "ring" links we mean intramolecular bonds formed by reactions such as



(ϵ represents the electron). Equation (3) is similar to a reaction proposed earlier.³ Other ring forming reactions could be proposed.

Cross links and vinylene groups formed by irradiation have been recognized since 1950,³ but ring or square links have never before been observed in the irradiation of any substance as far as we know. It is the purpose of this note to present evidence for cyclization during irradiation.

1,2-Disubstituted cyclopentane and cyclohexane derivatives absorb⁴ in the infrared approximately at wave lengths of 10.2 μ with extinction coefficients varying between 6 and 28 l. cm⁻¹. mole⁻¹. In Marlex-50 the initial absorption at 10.0 and 10.98 μ due to the vinyl group rapidly falls with irradiation at room temperature and *in vacuo* (initial $G(-\text{vinyl})$ equal to 7.7), revealing the growth of another absorption at 10.1 μ which is approximately the wave length at which *trans* 1,2-disubstituted cyclopentanes and hexanes absorb. We have attributed the growth of this peak to the formation of ring links of this size (other ring or square links which might absorb at this wave length are not excluded). A similar growth in the peak at 10.1 μ was found also in the case of a Ziegler-type polyethylene.

An order of magnitude estimate of G (*trans* ring links) is 1.0 which is of sufficient magnitude to show the necessity of considering cyclization in attempting to achieve a material balance.

In the case of a low density polyethylene, the vinylene peak at 10.3 μ is broader than in the case of Marlex-50 and possibly obscures the peak due to the ring systems. The 10.3 μ peak is unsymmetrical, however, which suggests that ring links are produced in low as well as high density polyethylenes.

This research was supported by a grant from the U.S. Atomic Energy Commission.

(3) M. Dole, Report of Symposium IV, "Chemistry and Physics of Radiation Dosimetry," Army Chemical Center, Md., September, 1950.

(4) "Catalog of Infrared Spectral Data" of the American Petroleum Institute Research Project, Carnegie Institute of Technology, Pittsburgh, Pennsylvania.

(5) Fulbright Fellow, 1956.

(6) On leave from A.E.R.E., Harwell, England.

DEPARTMENT OF CHEMISTRY
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RECEIVED JUNE 17, 1957

ENZYMATIC OXYGEN FIXATION INTO ACETATE CONCOMITANT WITH THE ENZYMATIC DECARBOXYLATION OF L-LACTATE

Sir:

The enzymatic incorporation of molecular oxygen into various organic substrates recently has been

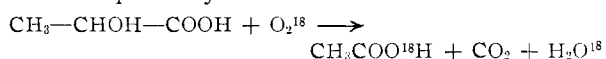
reported from several laboratories.¹ These "oxygenases"² are generally concerned with the transformation (hydroxylation, ring cleavage, cyclization) of various ring compounds such as aromatic amino acids, phenols and steroids. In this communication we wish to report a novel type of decarboxylation reaction in which atmospheric oxygen is incorporated into a simple aliphatic compound.

L-Lactate was incubated with a crystalline lactic oxidative decarboxylase prepared from *Mycobacterium phlei*³ in an atmosphere of O₂¹⁸ and with H₂O¹⁸ as a medium. Potassium acetate isolated from the incubation mixture was found to have incorporated approximately one atom of atmospheric oxygen (Table I). The O¹⁸ enrichment of the incorporated oxygen atom corresponded to approximately 82% of that of the atmospheric oxygen used in this experiment, assuming only one atom was inserted into the product. On the other hand, when the reaction was carried out in the medium of H₂O¹⁸ and with O₂¹⁶ as a gas phase, potassium acetate isolated did not contain appreciable O¹⁸. Carbon dioxide did not contain O¹⁸ in the first

case, but was highly enriched in the second experiment. This incorporation of O¹⁸ from H₂O¹⁸ into carbon dioxide probably was caused by a non-enzymatic exchange reaction as reported by Cohn and Urey⁴ and more recently by Rothberg and Steinberg in their studies of various microbial decarboxylases.⁵

The enzyme which catalyzes "oxidative" decarboxylation of L-lactate has been described from various microorganisms.⁶ The over-all reaction involves oxidation and decarboxylation of L-lactate to form stoichiometric quantities of acetate and carbon dioxide, but the mechanism of this unique enzymatic reaction has not yet been completely understood.

The available evidence suggests that at least one atom of atmospheric oxygen was incorporated into acetate when the C₁-C₂ bond of lactate was cleaved and that the other atom of the oxygen molecule probably was reduced to water



The mechanism of this reaction may be analogous to that of the so-called "mixed function oxygenase"^{1c,f} except that instead of reduced pyridine nucleotides, the substrate itself is providing electrons to reduce one atom of oxygen.

(4) M. Cohn and H. C. Urey, *THIS JOURNAL*, **60**, 679 (1938).

(5) S. Rothberg and D. Steinberg, *ibid.*, **79**, 3274 (1957).

(6) (a) N. L. Edson, *Biochem. J.*, **41**, 145 (1947); (b) N. L. Edson and F. B. Cousins, *Nature*, **171**, 702 (1953); (c) Y. Yamamura, M. Kusunose and E. Kusunose, *ibid.*, **170**, 207 (1952); *J. Biochem., Japan*, **39**, 227 (1952); (d) W. B. Sutton, *J. Biol. Chem.*, **210**, 309 (1954); **216**, 749 (1955).

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RECEIVED AUGUST 1, 1957

TABLE I
ENZYMATIC INCORPORATION OF O¹⁸ INTO ACETATE

Experiment I: Crystalline lactic oxidative decarboxylase³ (3,350 units), 2 millimoles of DL-lithium lactate and 2 millimoles of potassium phosphate buffer (pH 6.0) were incubated in a total volume of 50 ml. of water in a special flask designed for this type of experiment.^a An O₂¹⁸-helium gas mixture (2:3) was used as a gas phase.^b Experiment II: The same reaction components as in Experiment I were employed except that H₂O¹⁸ was used as a solvent and O₂¹⁶-helium gas mixture (2:3) was used as a gas phase. The incubation was carried out at 37° for 30 minutes with vigorous shaking. After the oxygen and carbon dioxide were removed for mass spectrometric analyses, the incubation mixture was immediately chilled at 4° and pH was adjusted to 3.5 with several drops of 2N H₂SO₄. The solution was then frozen and water and acetic acid were distilled from the frozen state. The distillate was neutralized with 1N KOH to pH 6.5 and water was removed by lyophilization. The residue dried over P₂O₅ was pyrolyzed^c and the O¹⁸ content determined with a mass spectrometer.^d

Expt.	Type	Atom % excess	Cpd. analyzed	Atom % excess found
I	O ₂ ¹⁸	6.31868	KOAc	2.59578
			CO ₂	0.00452
II	H ₂ O ¹⁸	1.3380	KOAc	0.01772
			CO ₂	1.32422

^a Details will be published elsewhere. See also 1b.
^b Highly enriched O₂¹⁸ gas was prepared by electrolysis of approximately 33% enriched H₂O¹⁸ purchased from the Weizmann Institute of Science, Israel. We are indebted to Dr. Y. Saito and Mr. L. Wartofsky for the preparation of O₂¹⁸ gas. ^c The mass spectrometric analyses were carried out in collaboration with Mr. W. E. Comstock of this Institute. ^d Pyrolysis was carried out at 400° for 1 hour with HgCl₂ as a catalyst according to D. Rittenberg and L. Ponticorvo (*Intern. J. Appl. Radiation and Isotopes*, **1**, 208 (1956)).

(1) (a) H. S. Mason, W. L. Fowles and E. Peterson, *THIS JOURNAL*, **77**, 2914 (1955); (b) O. Hayaishi, M. Katagiri and S. Rothberg, *ibid.*, **77**, 5450 (1955); (c) T. T. Tchen and K. Bloch, *ibid.*, **77**, 6085 (1955); (d) M. Hayano, M. C. Lindberg, R. I. Dorfman, J. E. H. Hancock and W. von E. Doering, *Arch. Biochem. and Biophys.*, **59**, 529 (1955). For general reference, see also recent reviews on this subject: (e) H. S. Mason, *Science*, **125**, 1185 (1957); (f) O. Hayaishi, Proceedings International Symposium on Enzyme Chemistry, in press.

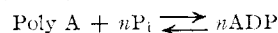
(2) O. Hayaishi, S. Rothberg and A. H. Mehler, Abstract of Papers, 130th American Chem. Soc. Meeting, Atlantic City, N. J., 1956. 53-C.

(3) W. B. Sutton, *J. Biol. Chem.*, **226**, 395 (1957).

POLYNUCLEOTIDE PHOSPHORYLASE IN LIVER NUCLEI

Sir:

Polynucleotide phosphorylase has been described in bacteria.¹ It catalyzes the reversible synthesis of polynucleotides from nucleoside diphosphates as well as the phosphorolysis of isolated ribonucleic acids. Thus, adenylic polynucleotide (Poly A) reacts with orthophosphate (P_i) to form adenosine diphosphate (ADP)



An enzyme has now been fractionated from guinea pig liver nuclei which converted up to 30% of the adenylic acid units of Poly A to ADP (Table I). Part of the ADP then formed ATP and AMP due to the presence of adenylate kinase. The rest of the polymer was utilized by a competing hydrolytic enzyme, shown to occur in liver nuclei.² ADP was identified by its R_f in four solvent systems, electro-

(1) (a) M. Grunberg-Manago, P. J. Ortiz and S. Oelha, *Biochim. et Biophys. Acta*, **20**, 269 (1956); (b) U. Z. Littauer, *Fed. Proc.*, **15**, 302 (1956); (c) R. F. Beers, Jr., *Nature*, **177**, 790 (1956).

(2) L. A. Heppel, P. J. Ortiz and S. Oelha, *Science*, **123**, 415 (1956).